

Antifungals in Clinical Use and the Pipeline



Melissa D. Johnson, PharmD, MHS

KEYWORDS

- Antifungal drugs • Polyenes • Azoles • Echinocandins • Pipeline
- Invasive fungal infections • Novel therapies • Review

KEY POINTS

- Polyenes, azoles, echinocandins, and the antimetabolite agent flucytosine are the most common antifungals currently used for treatment and prevention of systemic fungal infections.
- Current systemic antifungal therapy choices have unique profiles and challenges related to spectrum of activity, pharmacokinetic/pharmacodynamic properties, development of resistance, toxicities, and drug interactions.
- At least 7 new antifungals are in phase 2 or phase 3 clinical trials for invasive fungal infections and have potential advantages over older agents in terms of activity against resistant organisms, dosing/pharmacokinetics, and tolerability.
- This article reviews pharmacologic facets and clinical use of Food and Drug Administration-approved and pipeline antifungal agents.

INTRODUCTION

Over the past 15 years, there has been an increase in development and utilization of newer antifungal agents. The ideal antifungal agent in regard to spectrum of activity, pharmacokinetic/pharmacodynamic properties, development of resistance, safety, and drug interaction profile remains elusive. This article reviews pharmacologic aspects of currently available systemic antifungal agents in use for treatment and prevention of invasive fungal infections (IFIs). The discussion of pharmacokinetic and dosing parameters focuses on adults, but there are emerging data for children, reviewed comprehensively elsewhere.^{1,2} Promising antifungal candidates in clinical development also are discussed.

POLYENES

Amphotericin B (AmB) was the first antifungal approved for clinical use in systemic fungal infections and remains the sole polyene available today for this use. In addition

Duke University Medical Center, Box 102359 DUMC, Durham NC 27710, USA

E-mail address: johns200@mc.duke.edu

Infect Dis Clin N Am 35 (2021) 341–371

<https://doi.org/10.1016/j.idc.2021.03.005>

0891-5520/21/© 2021 Elsevier Inc. All rights reserved.

id.theclinics.com

to AmB deoxycholate (AmBd), there have been 3 Food and Drug Administration (FDA)-approved commercially available lipid formulations of AmB (LFABs): AmB lipid complex (ABLC), AmB colloidal dispersion, and liposomal AmB (LAmb). Only 2 of these lipid formulations (ABLC and LAmB), however, currently are marketed in the United States.

Mechanism of Action

Polyenes exert their antifungal activity by interacting with sterols in the fungal cell membrane, creating nonaqueous and aqueous channels that lead to leakage of cellular contents such as potassium through these pores.³ This leads to loss of membrane potential and subsequent cellular collapse.

Spectrum of Activity/Resistance

Polyenes have a broad spectrum of antifungal activity including yeasts, such as *Candida* spp, *Rhodotorula* spp, and *Cryptococcus* spp; molds, such as *Aspergillus* spp, *Rhizopus* spp, *Mucor* spp, *Rhizomucor* spp; and endemic fungi, such as *Histoplasma* spp, *Coccidioides* spp, and *Blastomyces* spp. AmB lacks activity, however, against a few fungi, including *Aspergillus terreus*, *C lusitaniae*, *C haemulonii* species complex, *Scedosporium apiospermum*, *Lomentospora prolificans*, and some *Fusarium* spp.⁴ Higher AmB minimum inhibitory concentrations (MICs) also have been reported for numerous strains of *C auris*.⁵ Alterations in fungal cell wall composition have been correlated with resistance to AmB in some species.^{6,7} A recent study of *C auris* also identified 4 single-nucleotide polymorphisms in protein-encoding regions that were associated with AmB resistance.⁸

Formulations/Pharmacokinetics

AmB and LFABs have poor oral bioavailability, so are typically administered intravenously (IV) for systemic infections. Each of these preparations has differential pharmacokinetic properties (Table 1). None of these formulations is metabolized by the cytochrome P450 (CYP) enzyme system; therefore, these agents have few drug-drug interactions (Table 2).

In addition to IV administration, AmBd or LFAB can be nebulized and delivered via inhalation. This frequently is performed in some centers as prophylaxis in high-risk patients, such as allogeneic hematopoietic stem cell transplant, and lung transplant, or as adjunctive therapy in difficult-to-treat pulmonary fungal infections.^{50–52}

Adverse Events

AmB has myriad associated adverse events that complicate IV administration, including infusion-related reactions (IRRs), nephrotoxicity, and electrolyte disturbances (hypokalemia/hyperkalemia and hypomagnesemia). IRRs have been reported in as many as 70% to 90% of patients receiving systemic AmBd.⁵³ Research over the past decade has illuminated the potential mechanism of AmBd-associated IRRs. Derived from *Streptomyces*, AmB is a ligand of Toll-like receptor 2 and CD14 on mononuclear cells, and, when recognized, initiates signaling pathways that lead to release of proinflammatory agents, such as interleukin (IL)-1b, IL-6, IL-8, and tumor necrosis factor α , that may be responsible for the IRRs that are observed.^{54,55}

AmB-associated nephrotoxicity likely is multifactorial. First, administration of AmB leads to vasoconstriction of the afferent renal arterioles, resulting in reduced renal blood flow and a drop in glomerular filtration.⁵⁴ In addition, binding of AmB to cholesterol in cell membranes of the renal tubules and uptake through low-density-lipoprotein-AmB complexes may lead to high concentrations of AmB and subsequent

Table 1
Key pharmacokinetic properties of approved antifungals and phases 2/3 pipeline agents

Drug Class/ Antifungal Agent	Status	Oral Bioavailability	Vss (L/kg)	AUC $\mu\text{g}(\text{0-24 h})$ $\mu\text{g}\cdot\text{h/mL}$	Peak Concentrations ($\mu\text{g/mL}$)	Protein Binding	P-gp Substrate/ Inhibitor	Metabolism/ CYP Inhibition	Half-Life (h) ^a
Polyenes									
AmBd ⁹⁻¹⁴	FDA approved	<5%	5.2	17.1	1.4	>95%	ND	Minor hepatic	6.8/127 ^b
LAmb ^{9,10,12}	FDA approved	<5%	0.1–0.4	171–1286	22.9	>95%	ND	None apparent	6.4–13.1/152 ^b
ABLC ^{9,10,15}	FDA approved	<5%	131	16.5–19.2	2.4–3.7	>95%	ND	None apparent	173.4–393
MAT2203 ¹⁶	Phase 2	ND	ND	2.0–2.2	0.06	ND	ND	ND	ND
Antimetabolite									
5-FC ^{10,17}	FDA approved	78%–89%	0.6	62	80	3%–4%	No	Minimal hepatic metabolism	2–5
Azoles									
Fluconazole ^{18,19}	FDA approved	95%	0.7	359–409	12.1–25	12%	Possible inhibitor	CYP2C19, CYP2C9, CYP3A4 inhibitor	30
Isavuconazole ^{20–22}	FDA approved	97%–98%	6	60–64	5–7.5	>99%	Weak inhibitor	CYP3A4, CYP3A5 substrate Moderate CYP3A4 inhibitor	110–115

(continued on next page)

**Table 1
(continued)**

Drug Class/ Antifungal Agent	Status	Oral Bioavailability	Vss (L/kg)	AUC $\mu\text{g}(\text{0-24 h})$ $\mu\text{g}\cdot\text{h/mL}$	Peak Concentrations ($\mu\text{g/mL}$)	Protein Binding	P-gp Substrate/ Inhibitor	Metabolism/ CYP Inhibition	Half-Life (h) ^a
Itraconazole ^{18,23}	FDA approved	55% ^c SUBA-itraconazole 81%	11	AUC(0- ∞) Capsule 14.9 SUBA-itraconazole 15.6	1.1-2	>99%	Substrate and inhibitor	CYP3A4 substrate CYP2C9, CYP3A4 inhibitor	34-42
Ketoconazole ²⁴⁻²⁶	FDA approved	84%	0.4	24	3.6	>93%	Inhibitor	CYP3A4 substrate and inhibitor	2-8
Posaconazole ^{18,27}	FDA approved	Susp: 8%-44% DR tab: 54% fasting (high fat meal ↑ 1.5 ×)	7-25	Susp: 8.5 DR tab 31.4 IV 42.9	Susp 0.6 DR tab 2 IV 2.6	>98%	Substrate	Glucur-onidation by UGT1A4 Strong CYP3A4 inhibitor	18.7-29.2
Voriconazole ^{18,28}	FDA approved	96%	4.6	AUC ₍₀₋₁₂₎ 34	4.6	58%	No	CYP2C19, CYP2C9, CYP3A4 substrate and inhibitor	6

Otseconazole ^{29–32}	Phase 3	73% ^d , ↑ with food	1.4 ^d	AUC _(0–8) 1.9–4.2 ^e	0.4–1 ^e	>99%	ND	Weak inhibitor CYP2C9, CYP2C19, CYP3A4	76–160 d
Glucan-synthase inhibitors									
Anidulafungin ^{19,33}	FDA approved	<5%	30–50L	55–93	3.9–7.7	>95%	No	Chemical degradation	24–26
Caspofungin ^{19,33,34}	FDA approved	<5%	9.5L ^f	52–170	3.9–13.8	>95%	No	Minor CYP3A4 substrate	9–11
Micafungin ³³	FDA approved	<5%	18L	158	0.24	>95%	No	Minor CYP3A4 substrate	10–17
Rezafungin ^{35–38}	Phase 3	<5%	28.5L	AUC _(0–168) 813	12.4 ^g	>98%	ND	Minimal P450 interaction	140
Ibrexafungerp ^{39–41}	Phase 3	>30%	>5	13.4	0.83	>99%	Substrate	CYP3A4 substrate CYP2C8 inhibitor	20–30
Orotomide									
Olorofim ^{42,43}	Phase 2	45%–82%	~3	35–55	4.6	>99%	ND	Multiple CYP enzymes CYP3A4 substrate Weak CYP3A4 inhibitor	20–30

(continued on next page)

Table 1
(continued)

Drug Class/ Antifungal Agent	Status	Oral Bioavailability	Vss (L/kg)	AUC _{0-24 h} μg·h/mL	Peak Concentrations (μg/mL)	Protein Binding	P-gp Substrate/ Inhibitor	Metabolism/ CYP Inhibition	Half-Life (h) ^a
Glycosylphosphatidylinositol inhibitor									
Fosmanogepix ^{44–46}	Phase 2	>90%	ND	315 ^b	21.3 ^b	ND	ND	Possible substrate of CYP2C19, CYP2D6, CYP3A4, and CYP3A5	52.6 ^b

Abbreviations: Vss, Volume of distribution at steady state; P-gp, P-glycoprotein; CYP, Cytochrome P450; Susp, suspension; [Up Arrow Symbol], increase; DR tab, delayed-release tablets; ND, has not been determined, still investigational.

^a Assuming normal organ function with standard dosing in adults.

^b Represents terminal half-life, due to triphasic elimination.

^c Bioavailability for capsules highest when taken immediately after a full meal; solution is 30% higher under fasted conditions.

^d Based on preclinical data in animal models.

^e After day 1 of dosing.

^f Likely larger than this due to limitations of experimental methods.

^g Day 8 of 200 mg daily.

^h Day 7 of 1000 mg daily.

Table 2

Pharmacodynamic characteristics and therapeutic drug monitoring recommendations for current and phases 2/3 pipeline antifungal agents^{47,48}

Drug Class or Investigational Agent	Concentration Dependent	Prolonged PAFE	PD Index Predictive of Efficacy	Serum Concentration Monitoring Recommended
Polyenes	Yes	Yes	Cmax/MIC	No
Antimetabolite	No	No	T > MIC	Yes
Azoles	No	Yes	AUC/MIC	Yes: voriconazole, itraconazole, posaconazole, possibly isavuconazole ND for oteseconazole
Echinocandins	Yes	Yes	Cmax/MIC or AUC/MIC	No
Ibrexafungerp ⁴⁰	Yes	ND	Cmax/MIC or AUC/MIC	ND
Fosmanogepix ⁴⁵	Yes	Yes	AUC/MIC	ND
Olorofim ⁴²	No	ND	Cmin/MIC ⁴⁹	Yes

Abbreviations: PAFE, Post Antifungal Effect; PD, Pharmacodynamic; Cmax, peak concentrations; Cmin, trough concentrations; DR tab, delayed-release tablets; ND, has not been determined, still investigational; T, time.

toxicity. Furthermore, higher proinflammatory cytokine/chemokine levels have been observed in patients with AmB-associated nephrotoxicity and may contribute to this phenomenon.⁵⁵

Nephrotoxicity may occur in a substantial number of patients receiving IV AmBd. In a recent Cochrane analysis of 10 randomized clinical trials, AmBd was associated with a greater than or equal to 2-fold increase in serum creatinine in approximately 26% of patients.⁵⁶ Saline loading prior to AmBd infusions has been proposed to help mitigate this toxicity, but there is a lack of data demonstrating clear benefit of this approach.^{53,54} Presence of factors, such as comorbid illnesses, receipt of concurrent nephrotoxins, AmB daily and total doses, and intensity of hospital care, have been associated with increased risk of AmBd-associated nephrotoxicity.⁵³

LFBAs generally have yielded lower rates of IRRs and nephrotoxicity than AmBd, because the lipid moiety prevents interaction of amphotericin with cholesterol in cell membranes and allows more selective and controlled release of drug over time. In the Cochrane analysis of 10 randomized clinical trials, LAmB was associated with a greater than or equal to 2-fold increase in serum creatinine in approximately 10% of patients, with an overall risk ratio of 0.49 (95% CI, 0.40–0.59) compared with AmBd.⁵⁶ In a broader recent systematic review and meta-analysis, including 5 different LFBAs compared with AmBd, LFBAs were found to be associated with lower IRRs (fever [odds ratio (OR) 0.49; 95% CI, 0.26–0.94]; chills [OR 0.44; 95% CI, 0.21–0.92]; vomiting [OR 0.64; 95% CI, 0.46–0.88], and nephrotoxicity [OR 0.32; 95% CI, 0.25–0.41]).⁵⁷

Clinical Use

AmB has a potential role in management of a wide variety of fungal infections, including candidiasis, cryptococcosis, aspergillosis, and mucormycosis. It also has been effective in endemic mycoses, such as histoplasmosis, coccidioidomycosis, blastomycosis, and sporotrichosis.⁵⁸ LFBAs often are substituted for AmBd for

many of these infections due to their improved tolerability.⁵⁹ LFABs are recommended by European Society for Clinical Microbiology and Infectious Diseases/European Confederation of Medical Mycology guidelines as options for treatment of fusariosis, invasive aspergillosis due to *Aspergillus calidoustus*, and invasive aspergillosis due to *Aspergillus* isolates with voriconazole MIC greater than or equal to 2.^{60,61} AmB also may have a role in treatment of infections due to *Acremonium* spp and *Purpureocillium lilacinum*, based on limited clinical data.⁶⁰

NOVEL POLYENE FORMULATIONS IN DEVELOPMENT

Other formulations have been extensively investigated to facilitate oral delivery of AmB with a favorable efficacy and safety profile.⁶² Of these, a cochleated form of AmB (MAT2203; Matinas BioPharma Nanotechnologies, Bedminster, NJ), is furthest along in clinical development. This agent currently is in phase 2, having completed a study in vulvovaginal candidiasis (VVC) (NCT02971007) with ongoing studies in mucocutaneous candidiasis (NCT02629419) and cryptococcal meningitis (NCT04031833). MAT2203 has been designated as a fast track, qualified infectious diseases product (QIDP) with orphan drug designation by the FDA for treatment of invasive candidiasis and aspergillosis, prevention of IFIs in patients receiving immunosuppressants, and treatment of cryptococcosis.

Cochleates are lipid delivery systems that have been investigated since the 1970s. MAT2203 is composed of a lipid bilayer rolled into a cigar-like spiral, encapsulating and protecting hydrophobic AmB in its interior.⁶³ Once in the body, cochleates fuse with target cell membranes and release AmB to the cytoplasm; MAT2203 also is thought to be readily taken up by macrophages, which facilitate delivery to sites of infection.⁶³ Experimental data from mice indicate that cochleated AmB is readily distributed into tissues with effective concentrations in the lungs, liver, kidneys, and spleen.^{64,65} MAT2203 also demonstrated measurable brain tissue concentrations after enteral administration in mice infected with *Cryptococcus neoformans*, although this was somewhat lower than that of AmBd.⁶⁶

In a small study evaluating MAT2203 for moderate to severe VVC, MAT2203 (100 mg or 200 mg orally, twice daily for 5 days) was compared with a single dose of fluconazole (150 mg orally) (NCT02971007). These results have not been published in a peer-reviewed journal, but clinical cure at day 12 reportedly was achieved in approximately 53% of MAT2203 recipients compared with 75% of fluconazole recipients.⁶⁷ No patients died or had a serious adverse event, and the most common adverse events included gastrointestinal disturbances, bacterial vaginosis, and urinary tract infections.

Phase 1 pharmacokinetic data recently were published and suggested similar exposures after MAT2203 doses of 1 g, 1.5 g, and 2 g daily, ranging from a median area under the concentration curve (AUC)_(0-24 h) of 1970 to 2180 ng·h/mL (see Table 1).¹⁶ Dividing the total daily dose into 4 to 6 divided doses in this study seemed to reduce some of the gastrointestinal disturbances compared with a previous MAT2203 phase 1 study where single doses of 800 mg were used. Based on these results, a total daily dose of 2 g was selected for induction therapy in combination with flucytosine (5-FC) followed by 1.5 g daily in divided doses plus fluconazole for cryptococcal meningitis.¹⁶

Additional studies are needed to fully evaluate optimal dosing, efficacy, and safety of MAT2203 but this is a promising option for oral polyenes in the treatment or prevention of systemic fungal infections, such as candidiasis, aspergillosis, and cryptococcosis.

ANTIMETABOLITE

The antimetabolite antifungal 5-FC was approved by the FDA in the 1970s and is used primarily as adjunctive therapy for a variety of fungal infections.

Mechanism of Action

The antimetabolite 5-FC exerts its effect inside fungal cells after conversion to its active form, 5-fluorouracil, by fungal specific cytosine deaminase. It then disrupts several cellular processes, including RNA, protein, and DNA synthesis.

Spectrum of Activity/Resistance

Cryptococcus spp, *Aspergillus* spp, some dematiaceous fungi, and many *Candida* spp (except *C krusei*) are susceptible to 5-FC.^{17,68,69} Fungal resistance to 5-FC can be intrinsic or acquired. Given concerns about resistance, 5-FC is combined most often with other antifungals for treatment of IFIs.

Formulations/Pharmacokinetics

5-FC is administered orally in the United States and has high bioavailability (see **Table 1**). 5-FC distributes widely into tissues, including cerebrospinal fluid (CSF). It is eliminated renally and requires dose adjustment in those with renal impairment.¹⁷

Adverse Events

Most commonly, 5-FC side effects include gastrointestinal disturbances and hepatic effects, including elevations in liver enzymes. Bone marrow suppression can also occur, especially with 5-FC concentrations above 100 mg/L.⁷⁰ Coadministration with AmB can lead to reduced excretion of 5-FC in patients who experience AmB-induced nephrotoxicity, leading to higher 5-FC concentrations. Coadministration with other myelosuppressive agents can be problematic and compound bone marrow effects of 5-FC.⁷⁰

Clinical Use

The antimetabolite 5-FC is first line in combination with AmB for treatment of cryptococcal meningitis and severe cryptococcal pneumonia.⁷¹ It also may be used adjunctive therapy (mostly combined with AmB formulations) for difficult-to-treat invasive *Candida* spp infections and alone or in combination for certain clinical cases of fluconazole-resistant *C glabrata* cystitis/pyelonephritis.⁷² 5-FC was used historically for some cases of chromoblastomycosis, but its use has been supplanted by more effective agents, such as itraconazole and terbinafine.⁷³ It still may be useful in combination with other antifungals for mycetoma and cerebral infections with certain dematiaceous fungi, such as *Cladophialophora*.⁷⁴

AZOLES

Triazoles are the most common antifungal class administered systemically in the United States and have 3 nitrogens on their azole ring. This imparts increased enzyme specificity and a better safety profile than most imidazoles, which typically are reserved for topical administration. There is 1 imidazole that is FDA approved for systemic use, ketoconazole, as well as 5 triazoles: fluconazole, itraconazole, voriconazole, posaconazole, and isavuconazole.

Mechanism of Action

Azoles inhibit ergosterol formation in fungal cell membranes by targeting the enzyme CYP-dependent 14α -demethylase (CYP51), which converts lanosterol to ergosterol. This results in accumulation of methylsterols, disruption of fungal cell membranes, and inhibition of cellular growth and replication.⁵³

Spectrum of Activity/Resistance

Differences in chemical structure and affinity for sterol 14α -demethylase imparts differential antifungal activity between the azoles. Fluconazole is active against most *Candida* and *Cryptococcus* spp, whereas ketoconazole, itraconazole, voriconazole, posaconazole, and isavuconazole have activity against yeasts and molds.

Intrinsic resistance to some azoles, such as fluconazole, may occur naturally due to amino acid substitutions in sterol 14α -demethylase encoded by the gene CYP51A (ERG11).⁷⁵ Such mutations also may develop within a formerly susceptible species and lead to decreased affinity for binding of the azole to the enzyme. Several other mechanisms of azole resistance have been reported, including up-regulated ERG11 expression, leading to overproduction of sterol 14α -demethylase or overexpression of efflux pumps.⁷⁶ Increasing resistance among formerly susceptible *Candida* spp and *Aspergillus* spp has been noted and occurs more often in settings where previous azole exposure has occurred through therapeutic or environmental applications.^{77–82}

Formulations/Pharmacokinetics

Many azoles are available in formulations for oral as well as IV delivery, which allow versatile dosing options in a variety of conditions. There is no IV form of ketoconazole, however, and the IV form of itraconazole has been withdrawn from the US market. Each of the azoles has distinct pharmacokinetic properties (see Table 1), and there have been notable challenges in achieving reliable systemic concentrations with some of these agents/formulations.

Therapeutic drug monitoring (TDM) has been proposed for many of the mold-active azoles, including itraconazole, voriconazole, and posaconazole (see Table 2).^{47,83} TDM for these azoles may be especially relevant in patients with hypermetabolic conditions, drug-drug interactions, treatment failure, conditions that increase risk of poor absorption when receiving oral azoles, or if toxicity is suspected.^{61,83} Additional data are needed to fully establish exposure-response relationships with isavuconazole, but it appears that patients in initial clinical trials achieved effective plasma concentrations with standard dosing.²⁰ Newer data suggest there may be a role of TDM for isavuconazole in certain clinical situations.^{61,84–86}

Itraconazole and its active metabolite, hydroxyitraconazole, have nonlinear and variable pharmacokinetics. Dissolution of the drug is pH dependent and requires an acidic environment for adequate absorption, especially in capsule form. Newer oral formulations have been developed to try to overcome these pharmacokinetic limitations, including a cyclodextrin suspension and more recently superbioavailability itraconazole (SUBA-itraconazole [Tolsura], Mayne Pharmaceuticals, Raleigh, NC). SUBA-itraconazole capsules contain a pH-dependent polymer matrix with a solid dispersion of itraconazole. In an analysis of 7 studies, SUBA-itraconazole showed enhanced bioavailability (173% [95% CI, 156%–190%]) and lower variability (by 21%) in absorption compared with itraconazole capsules.⁸⁷ Pharmacokinetic models suggested 58 mg of SUBA-itraconazole yielded similar concentrations as 100 mg of itraconazole capsules. Food appeared to reduce absorption of both SUBA-itraconazole and

itraconazole capsules similarly, with a reduction of approximately 20% in AUC and 35% in peak serum concentrations.⁸⁷

Voriconazole is known to have substantial intrapatient and interpatient pharmacokinetic variability, with nonlinear pharmacokinetics. Voriconazole has high bioavailability (96%) and moderate protein binding (58%), with CSF/plasma concentration ratios of 0.22 to 1.0.¹⁸ It is metabolized by CYP2C19, CYP3A4, and more minimally CYP2C9. Genetic polymorphisms in CYP2C19, and possibly CYP3A4, have been shown to have an impact on voriconazole pharmacokinetics.⁸⁸ Five metabolizer phenotypes of CYP2C19 now have been identified, ranging from poor to ultrarapid metabolizers.⁸⁹ Poor and heterozygous extensive CYP2C19 metabolizers have 4-fold and 2-fold higher concentrations than homozygous extensive metabolizers, respectively⁹⁰; 15% to 20% of Asian populations and 3% to 7% of whites and blacks are poor metabolizers, whereas 1% of Asians and more than 30% of whites and blacks may be ultrarapid metabolizers.^{90,91} Recommendations from the Clinical Pharmacogenetics Implementation Consortium (2017)⁸⁹ suggest use of an antifungal other than voriconazole in patients who are rapid and ultrarapid CYP2C19 metabolizers since attaining target concentrations with standard dosing is unlikely. Use of alternative antifungals for primary therapy or voriconazole dose reduction with TDM is recommended for poor CYP2C19 metabolizers, because higher concentrations are likely in this population and may increase the risk of adverse events.⁸⁹ Altered fluconazole and voriconazole pharmacokinetics have been reported in patients on extracorporeal membrane oxygenation (ECMO), due to drug sequestration within the circuit.⁹² In addition, dosing of voriconazole for obese patients based on total body weight has been associated with higher concentrations in some studies.^{93,94} TDM, therefore, also is recommended for voriconazole in patients on ECMO or with obesity.

Posaconazole has 3 available formulations: oral suspension, delayed release tablets, and solution for IV injection. Posaconazole oral suspension displays saturable absorption, whereby doses above 800 mg are not absorbed readily.⁹⁵ Overall concentrations were 2.5-fold higher when posaconazole suspension was administered as 200 mg 4 times daily compared with 400 mg twice daily.⁹⁶ The delayed-release tablets of posaconazole have overcome some of the limitations of its oral suspension, providing better absorption and higher concentrations.⁹⁷ For this reason, dosing of oral posaconazole is formulation-dependent, and careful attention to formulation must be considered when making dosing recommendations for this agent.

Isavuconazole is available in both IV and oral formulations as the more water-soluble prodrug, isavuconazonium acetate, which is rapidly converted by plasma esterases to active isavuconazole.⁹⁸ It exhibits dose-dependent kinetics, with wide tissue distribution. In studies to date, there has been limited interpatient variability reported, although altered concentrations have been reported in those receiving renal replacement therapy or ECMO/Cytosorb (Monmouth Junction, NJ) adsorber therapy.⁸⁴ Low concentrations also have been reported in CSF or brain tissue in the presence of uninflamed meninges, with higher concentrations in infected brain tissue or in the presence of inflammation.⁸⁵

All of the currently marketed azoles are substrates and/or inhibitors of CYP isoenzymes. Some also are inhibitors of P-glycoprotein (see Table 1). Together these features lead to substantial and unique drug interactions profiles of each azole that must be considered.⁹⁹ In a large study evaluating data from 150 hospitals, at least 88% of patients receiving mold-active azoles were administered at least 1 interacting concomitant drug.¹⁰⁰ Some of the more frequent interactions in clinical practice occur with antinausea medications, proton pump inhibitors, fentanyl, midazolam, immunosuppressants, and oral anticoagulants. Other interacting drugs include statins,

amiodarone, vincristine, venetoclax, midostaurin, oral anticoagulants, and anticonvulsants.^{100,101}

Safety

Cross-inhibition of some CYP-dependent enzymes in humans by azoles leads to some toxicities and drug interactions. Adverse effects common to azoles as a class include QTc prolongation, with torsades de pointes and hepatic enzyme elevations/hepatotoxicity.⁵³ Isavuconazole, however, has been reported to shorten QTc interval, so does not follow this class effect.¹⁰² Some adverse effects may be even more apparent with long-term azole exposure, such as alopecia, peripheral neuropathy, and hormone-related adverse effects.¹⁰³ Azoles in current clinical use are not recommended for use in pregnant women due to concerns about teratogenicity.^{104–107}

Most azoles also are known to cause gastrointestinal side effects, such as nausea and diarrhea. Itraconazole also has been associated with rash, hypokalemia, headache, fever, and dizziness. In addition to these effects, voriconazole-related photopsia occurs in approximately 20% to 30% of patients; this effect seems related to the level of serum concentrations and typically subsides in the first week of therapy.⁵³ Visual hallucinations and central nervous system (CNS) effects also have been reported with voriconazole, especially at higher serum concentrations.^{108,109} Skin rash, photosensitivity, and Stevens-Johnson syndrome also have been reported with voriconazole. More recently, an increase in squamous cell carcinomas has been observed in patients receiving voriconazole, especially with long-term exposure.¹¹⁰ Periostitis and exostoses have been reported in patients receiving voriconazole and may relate to accumulation of fluoride over long periods of voriconazole intake and osteogenic impact of voriconazole on osteoblasts. Fluconazole, posaconazole, and isavuconazole contain a lower amount of fluoride than voriconazole, and these side effects have not been reported with these other azoles to date.¹⁰³

Hormone-related changes, derangements in electrolytes, and adrenal suppression are variable within the azole class, due to variations in binding affinity for CYP-dependent enzyme CYP51A1, which converts lanosterol to ergosterol in the steroidogenesis pathway. Ketoconazole, and to a lesser extent itraconazole, blocks several other enzyme target sites in the steroidogenesis pathway, having an impact on formation of testosterone and estradiol and leading to side effects, such as gynecomastia, impotence, decreased libido, oligospermia, and azoospermia.¹⁰³ Adrenal insufficiency has been reported with all of these azoles with the exception of isavuconazole to date; pseudohyperaldosteronism also has been reported with itraconazole and posaconazole.^{103,111,112} Mineralocorticoid excess as well as potential direct damage to myofibroblasts or mitochondrial dysfunction with itraconazole may lead to cardiac toxicity and congestive heart failure with its use in some patients, but this does not appear to be a azole class effect.^{113,114}

IV formulations of voriconazole and posaconazole contain the vehicle sulfobutyl- β -cyclodextrin (SBECD), which may accumulate in patients with renal impairment (estimated glomerular filtration rate <50 mL/min).^{115,116} There have been concerns about necrosis of the liver and dose-dependent changes in renal histopathology, including vacuolation, in rats at high doses over prolonged exposure periods (1–6 months).¹¹⁷ Thus, the labeling for these formulations states they should be used only in patients with renal impairment if the benefit outweighs the risk. More recent reviews and retrospective analyses have suggested that there is not strong evidence demonstrating worsening renal dysfunction in patients with renal impairment who received IV voriconazole, and SBECD may be removed by dialysis.^{118,119} Itraconazole oral solution is solubilized with hydroxypropyl- β -cyclodextrin (HP- β -CD), which has

been associated with pancreatic adenocarcinomas in rat but not murine carcinogenicity studies. Toxicity of HP- β -CD at the concentrations present in itraconazole oral solution has not been apparent in humans to date.^{120,121}

Clinical Use

Given their versatility, azoles are the most prescribed class of systemic antifungal agents in United States hospitals.¹²² Fluconazole accounted for more than 80% of inpatient antifungal administrations from 2006 to 2012, with voriconazole comprising 6%.¹²² Fluconazole may be used in certain clinical situations for the treatment or prevention of candidemia and invasive candidiasis, coccidioidomycosis, and less severe cryptococcosis or as consolidation therapy following induction with LAmB and 5-FC for cryptococcal meningitis. Itraconazole maintains a niche primarily in treating certain cases of histoplasmosis, sporotrichosis, coccidioidomycosis, and non-CNS blastomycosis.^{123–126} Voriconazole is a first-line agent for invasive aspergillosis and also may be useful for fusariosis, *Scedosporium apiospermum* infections, certain clinical situations in *L. prolificans* and other rare mold infections, candidemia/invasive candidiasis, and cryptococcosis.^{60,71,72,124,127} Posaconazole is a first-line agent for prevention of fungal infections in patients with hematologic malignancies as well as treatment of some cases of aspergillosis, fusariosis, mucormycosis, histoplasmosis, blastomycosis, cryptococcosis, candidiasis, coccidioidomycosis, and certain other rare mold infections.^{60,61,71,72,124,125,127–131} Isavuconazole has a first-line role for invasive aspergillosis, and there are evolving data to support use in mucormycosis as well as certain clinical scenarios with candidiasis, cryptococcosis and dimorphic fungi.^{61,128,132}

NEW AZOLES IN DEVELOPMENT

In recent years, there have been at least 3 new tetrazole-based compounds in development: oteseconazole (formerly VT-1161), VT-1598, and quilseconazole (formerly VT-1129). These agents are highly selective for lanosterol 14 α -demethylase, yielding potent antifungal activity with the potential for fewer off-target effects leading to toxicity and drug interactions.¹³³

Oteseconazole and quilseconazole (Mycovia Pharmaceuticals, Durham, NC) have shown in vitro activity against fluconazole-resistant *C. krusei* and fluconazole and echinocandin-resistant *C. glabrata* at clinically achievable concentrations, suggesting promise for more difficult to treat candida infections.¹³⁴ Oteseconazole (Mycovia Pharmaceuticals, Durham, NC) is being studied in phase 2 and phase 3 trials for acute/recurrent VVC (NCT02267382, NCT01891331, NCT03561701, NCT03562156, and NCT01891331) and has additional phase 2 data evaluating it in the treatment of onychomycosis (NCT01891305) and tinea pedis (NCT01891305). Oteseconazole has been awarded QIDP status and a fast track designation for recurrent VVC by the FDA.¹³⁵ Phase 3 trials for recurrent VVC recently have been completed, and a New Drug Application (NDA) for this indication is expected to be filed in 2021.

In a phase 2 study of symptomatic acute VVC, oteseconazole oral doses of 300 mg daily, 600 mg daily, and 600 mg twice daily for 3 days were compared with fluconazole, 150 mg, as a single dose.²⁹ In intention-to-treat analysis, clinical and microbiologic cure at 28 days was similar for oteseconazole compared with fluconazole: 64.3% (300 mg daily), 75% (600 mg daily), and 78.6% (600 mg twice daily) versus 66.7%. With oteseconazole's long half-life, measurable drug concentrations were still observed at 168 days (127 ng/mL, 409 ng/mL, and 1300 ng/mL for initial 3-day courses of 300 mg daily, 600 mg daily, and 600 mg twice daily, respectively). No

patients treated with this new agent experienced recurrent VVC whereas 46% of fluconazole-treated patients had mycologically proved recurrence after 6 months.

VT-1598 (Mycovia Pharmaceuticals) currently is being evaluated in a phase 1 pharmacokinetic study and has shown activity against a range of fungi, such as *Candida* spp (including *C auris*), *Cryptococcus* spp, *Aspergillus* spp, *Rhizopus arrhizus*, and endemic fungi including *B dermatitidis*, *Coccidioides* spp, and *H capsulatum*.^{136,137} It has been granted QIDP status and a fast track as well as orphan drug designation for coccidioidomycosis by the FDA.

Quilseconazole demonstrated efficacy with linear pharmacokinetics after oral dosing for cryptococcal meningitis in immunocompetent mice.¹³⁸ This agent has a long half-life of more than 6 days in mice and displayed excellent CNS penetration in this study. Quilseconazole has been identified as a QIDP and received fast track and orphan drug designations from the FDA for treatment of cryptococcal meningitis. At this time, however, there are no quilseconazole clinical trials registered in the United States.

ECHINOCANDINS

Echinocandins are the newest antifungal class approved for use and include anidulafungin, caspofungin, and micafungin. These compounds consist of an amphiphilic cyclic hexapeptide with an *N*-linked acyl lipid side chain. Differences in the side chain make each echinocandin structurally unique.⁵³

Mechanism of Action

Echinocandins exert their antifungal effects by noncompetitively inhibiting the synthesis of (1 → 3)-β-D-glucan, an essential component of the fungal cell wall. They exert this effect by targeting the catalytic subunit of the enzyme complex (1 → 3)-β-D-glucan synthase, which is encoded by FKS genes.¹³⁹ Decreased production of (1 → 3)-β-D-glucan leads to a loss of cell wall integrity and subsequently, lysis of the cell.

Spectrum of Activity/Resistance

Echinocandins have fungicidal activity against *Candida* spp as well as fungistatic activity against *Aspergillus* spp. Echinocandins lack appreciable clinical activity against *Cryptococcus* spp, zygomycetes, *Fusarium solani*, and *Trichosporon* spp.^{140,141} Mutations in FKS1 and FKS2 in *Candida* spp have been associated with higher echinocandin MICs and diminished clinical activity.^{142–144} Echinocandins have variable activity against endemic fungi, with micafungin having the most potent activity in vitro against mycelial forms of *H capsulatum*, *B dermatitidis*, and *Coccidioides immitis* but little activity against yeast-like forms of these fungi. Echinocandins also have activity against the cyst forms of *P jirovecii* but not the trophozoite form.¹⁴¹

Formulations/Pharmacokinetics

Currently licensed echinocandins are available only in IV formulations. These compounds are large and do not penetrate CNS well. All are typically dosed once daily, but the half-life of these agents varies from 9 hours to 11 hours for caspofungin and 24 hours to 26 hours for anidulafungin. Echinocandins are not metabolized via the CYP system, so they have fewer drug interactions than azole compounds.¹⁴¹ In addition, their large size and high protein binding seem to limit removal of drug by hemodialysis or hemofiltration, although adsorption of micafungin to polyacrylonitrile (AN-69) membranes during continuous venovenous hemodiafiltration has been associated with lower exposures in some patients.¹⁴⁵ Dose modifications for patients

receiving echinocandins while undergoing hemodialysis or hemofiltration are not universally recommended.¹⁴⁶ Recent data suggest, however, altered pharmacokinetics of micafungin but not caspofungin or anidulafungin in patients on ECMO.^{146–150} Higher micafungin doses (at least 200 mg daily for adults) may be needed in these patients.

In addition, obese patients may experience inadequate target attainment due to lower-than-expected drug exposures with standard dosing of all echinocandins. Therefore, dose increases of these agents are recommended in obesity.^{151–154} Although original product labeling recommended maintenance dose reduction of caspofungin to 35 mg in patients with moderate hepatic insufficiency, a more recent study in patients with decompensated Child-Pugh class B or class C cirrhosis suggested this would result in lower than comparable exposures in noncirrhotic patients and potentially suboptimal concentrations.¹⁵⁵

Safety

Echinocandins have a favorable safety profile and are well tolerated by most patients. There is no synthesis of (1 → 3)- β -D-glucan in human cells as in fungal cell walls; thus, off-target effects of echinocandins in humans are less likely to occur. The most common adverse effects noted with echinocandins in clinical trials have been elevations in liver enzymes, gastrointestinal disturbances, hypokalemia, fever, and injection site reactions.^{156–158} In preclinical toxicology studies of long-term high-dose micafungin exposure, liver tumors were observed in rats, leading to a black box warning in European product labeling.¹⁵⁹ An evaluation of more than 23,000 patients who received systemic antifungal therapy in US hospitals, however, found no signal of hepatocellular cancers with micafungin compared with other antifungals in this population after 71,285 person-years of follow-up.¹⁶⁰ Short-term risk of liver injury and kidney injury also was found to be similar for patients receiving micafungin and other systemic antifungals in a propensity score matched analysis.¹⁶¹

Clinical Uses

Echinocandins are considered first-line agents for the treatment of candidemia in neutropenic and non-neutropenic adults and have a role in certain other forms of invasive candidiasis, such as intra-abdominal infections, osteomyelitis/septic arthritis, esophagitis, and chronic disseminated candidiasis.⁷² High-dose echinocandin therapy may be used for some cases of endocarditis, cardiac device infections, or suppurative thrombophlebitis due to susceptible *Candida* spp. Another role for echinocandins is salvage therapy or as part of combination therapy for invasive aspergillosis. Micafungin also has been shown to be noninferior to fluconazole as prophylaxis in hematopoietic stem cell transplant recipients in a randomized trial, and caspofungin demonstrated efficacy and better tolerability than LAmB in a randomized trial as empirical therapy of neutropenic fever.^{162,163}

NEW AGENTS IN DEVELOPMENT TARGETING GLUCAN SYNTHESIS

Rezafungin

Rezafungin (CD101, Cidara Therapeutics, San Diego, CA) is a next-generation echinocandin that is a structural analog of anidulafungin, with the same side chain with a slight change in the cyclic core, which yields improved stability and solubility. Rezafungin has a much longer half-life (>130 hours) than precursor echinocandins, which facilitates less frequent dosing. It also exhibits dose-proportional pharmacokinetics with little interpatient variability and a favorable safety profile, including lack of apparent effects on QT interval at doses up to 1400 mg.^{35,164} This agent has a

QIDP and fast track designation for prevention of IFIs in adults undergoing allogeneic bone marrow transplantation. It also is designated as a QIDP, fast track, and orphan drug designation for the treatment of candidemia/invasive candidiasis. Current phase 3 clinical trials are exploring rezafungin for both of these indications. In a phase 3 candidemia/invasive trial (ReSTORE, NCT03667690), rezafungin is administered as a 400-mg IV loading dose in week 1, followed by 200-mg IV weekly thereafter, and compared with daily caspofungin infusions with stepdown to oral daily fluconazole dosing. In the phase 3 prophylaxis trial (ReSPECT, NCT04368559), rezafungin is dosed similarly and compared with standard doses of trimethoprim/sulfamethoxazole (for *Pneumocystis jirovecii* prophylaxis) in combination with either fluconazole or posaconazole. Dosing in these trials was selected on the basis of prior dose-ranging studies and a phase 2 clinical trial evaluating rezafungin for candidemia/invasive candidiasis.

In a phase 2 randomized, double-blind multinational trial for candidemia and invasive candidiasis (STRIVE), rezafungin was compared with caspofungin (70 mg IV loading dose followed by 50 mg IV daily, with optional stepdown to oral fluconazole).³⁶ Rezafungin had 2 dosing schemes: either 400 mg IV weekly or 400 mg IV once during the first week followed by 200 mg IV weekly, for 2 weeks to 4 weeks total. Overall response in the modified intention-to-treat analysis at day 14 was 60.5% for rezafungin, 400 mg weekly; 76.1% for rezafungin, 400/200 mg weekly; and 67.2% for caspofungin; 30-day all-cause mortality rates were 15.8%, 4.4%, and 13.1% for these groups, respectively. There were several patients in the rezafungin 400-mg weekly arm with indeterminate responses at day 14 (13.2%, due to lack of data from mycologic cultures or assessment of systemic signs of infection), contributing to the trend in lower success rates in this group. These differences in outcome were apparent in mycological and clinical cure rates at day 5, after only a single 400-mg dose in each of the rezafungin groups. Median time to blood culture negativity was lower overall for patients receiving rezafungin (19.5 h) versus caspofungin (22.8 h; $P = .02$). The most common treatment emergent adverse events were hypokalemia (16.4%, rezafungin, and 13.2%, caspofungin), gastrointestinal disturbances, fever, and anemia. The rates of severe and serious treatment-related adverse events were similar between groups. Together these data support the comparative safety of rezafungin and proof-of-concept for efficacy with a front-loaded weekly dosing strategy in treating candidemia/invasive candidiasis.

Rezafungin was also previously evaluated as a topical treatment (as 3% gel or 6% ointment) for acute VVC in a phase 2 trial (NCT02733432)¹⁶⁵; 90% of participating women had *C albicans* infections, and 36% had a history of recurrent VVC. Although there was a relatively small number of nonalbicans isolates in this study, failure of nonalbicans infections was relatively high across all treatment groups (75%, rezafungin, and 100%, fluconazole). Overall, clinical cure rates were no better for either of the 2 rezafungin topical formulations than a single (150-mg) dose of fluconazole. Thus, topical rezafungin is not currently being developed further for the VVC indication.¹⁶⁶

Ibrexafungerp

Ibrexafungerp (SCY-078, Scynexis Pharmaceuticals, Jersey City, NJ) is a glucan synthase inhibitor with a unique chemical structure as a triterpenoid derivative of enfumafungin. Although it is not an echinocandin, it inhibits glucan synthesis through interaction with enzyme complex (1 → 3)- β -D-glucan synthase at a site that is distinct but partially shared with echinocandins.¹³⁹ This leads to some potential cross-resistance with echinocandins; however, studies to date suggest that ibrexafungerp retains activity against many *Candida* isolates with FKS mutations associated with

echinocandin-resistance.¹⁶⁷ It also appears active against many *C. auris* strains, including isolates that may be resistant to echinocandins and azoles.¹⁶⁸ Ibrexafungerp also has demonstrated in vitro activity against *Aspergillus* spp as well as *Paecilomyces variotii* and otherwise pan-resistant *L. prolificans*.¹⁶⁹

Ibrexafungerp is being studied in current clinical trials as an oral formulation. A liposomal formulation for IV administration is under development with plans for a phase 1 pharmacokinetic study in 2021.¹⁷⁰ This compound is widely distributed in tissues, including abscesses in experimental models, although it has relatively poor CNS penetration.^{171,172} It exhibits linear pharmacokinetics with a terminal half-life of 20 hours to 30 hours.³⁹ It is a substrate of CYP3A4, and ibrexafungerp dosing may need adjusted when coadministered with potent CYP3A4 inhibitors.³⁹ Although in vitro data have suggested that ibrexafungerp is a possible inhibitor of CYP2C8, it did not appear to significantly impact concentrations of rosiglitazone (a CYP2C8 substrate) in a clinical pharmacokinetics study.¹⁷³

Completed and ongoing clinical trials have focused on evaluating ibrexafungerp for the treatment of acute/recurrent VVC (NCT03987620, NCT03734991, NCT04029116, NCT02679456, and NCT03253094), *C. auris* infections (NCT03363841), candidemia/invasive candidiasis after initial therapy with an echinocandin (NCT02244606), a variety of fungal infections refractory/intolerant to other antifungals (NCT03059992), and invasive pulmonary aspergillosis in combination with voriconazole (NCT03672292).

Initial results with ibrexafungerp for VVC have been promising but are not yet published in peer-reviewed journals. A dose of 300 mg, twice daily for 1 day, was studied in 2 phase 3 trials for acute VVC and was significantly more effective than placebo for both clinical and mycological response.³⁹

In invasive candidiasis, results of a small ($N = 27$), phase 2, open-label study of step-down therapy after echinocandins indicated that ibrexafungerp had a similar response as standard of care (SOC), which was fluconazole step-down or micafungin for fluconazole-resistant isolates.¹⁷⁴ Ibrexafungerp had 2 dose groups: 1000-mg loading dose, followed by 50 mg daily, or 1250-mg loading dose, followed by 1000 mg daily, whereas fluconazole was given as an 800-mg loading dose, followed by 400 mg daily. Only 1 patient in the SOC arm received micafungin (100 mg daily) instead of fluconazole due to an azole-resistant *Candida* spp. Global responses at the end of therapy were 71% and 85% for ibrexafungerp maintenance doses of 500 mg and 750 mg, respectively, and 71% for SOC. The treatments were well tolerated, with the most common side effects of gastrointestinal disturbances (43%), infection/infestation (38%), and headache (24%). In a pharmacokinetic model, 85% of ibrexafungerp patients were found to achieve target steady-state AUC_{0-24} exposures ($15.4 \mu\text{M}\cdot\text{h}$, equivalent to $11.2 \mu\text{g}\cdot\text{h}/\text{ml}$).⁴⁰ One patient in the ibrexafungerp, 500-mg, maintenance dose group had suboptimal concentrations below the organism MIC but still responded to therapy. Only 1 patient in either ibrexafungerp group achieved the target AUC_{0-24} on day 1 of dosing.¹⁷⁴

The FURI study (NCT03059992) extends ibrexafungerp experience to infections refractory to or intolerant of other antifungals. Preliminary results of this open-label phase 3 study have suggested promising ibrexafungerp activity against a variety of deeply invasive *Candida* spp infections as well as some cases of mucocutaneous candidiasis.¹⁷⁵ In this study, ibrexafungerp is administered orally, at a dose of 750 mg twice daily for 2 days, followed by 750 mg daily for IFIs, with a dose reduction to 500 mg daily when combined with mold-active azoles.

Ibrexafungerp has been designated as a QIDP with fast track status by the FDA for treatment of VVC and prevention of recurrent VVC, and in 2020 an NDA was filed for the treatment of VVC with plans for priority review in 2021. It also has been designated

an orphan drug with QIDP and fast track status for oral use in treating invasive candidiasis, including candidemia, and invasive aspergillosis.¹⁷⁶

OTHER NOVEL AGENTS IN THE PIPELINE (PHASE 2 OR PHASE 3)

Orotomides

Olorofim (F901318, F2G, Manchester, UK) is the first of a novel antifungal class, the orotomides. These agents inhibit dihydroorotate dehydrogenase (DHODH) in the pyrimidine biosynthesis pathway, which is an essential component of many cellular processes, including nucleic acid, cell wall, and phospholipid synthesis.¹⁷⁷ Fungal cell regulation and protein production are also affected. Olorofim leads to fungal cell lysis and lack of fungal cell viability by reducing available substrates for formation of the cell wall components chitin and (1 → 3)- β -D-glucan as well as impacting hyphal growth, septation, and vacuole formation.^{42,178} Toxicity in humans is expected to be low, because olorofim's affinity for fungal DHODH is more than 2200-times greater than that of the human enzyme homolog.¹⁷⁷

In *in vitro* studies, olorofim has demonstrated activity against *Aspergillus* spp, *Scedosporium* spp, *L. prolificans*, *Penicillium*, *Paecilomyces*, *Purpureocillium*, *Talaromyces*, and *Microascus/Scopulariopsis* spp; and endemic fungi, such as *Coccidioides* spp, *Histoplasma* spp, and *Blastomyces* spp. It also has activity against some *Fusarium* spp but lacks activity against *Candida* spp, *Cryptococcus* spp, and *Mucorales*.^{42,179}

Clinical studies to date have focused on establishing the pharmacokinetic (see **Table 1**) and drug-interaction profile of olorofim in multiple formulations. To solubilize the drug, early IV formulations contained a HP- β -CD vehicle and oral tablets were formulated with hydroxypropyl methylcellulose acetate succinate.⁴² Oral bioavailability of 45% to 82% has been reported as well as enterohepatic recirculation.¹⁸⁰ This compound appears to be widely distributed, with some CNS penetration. It is metabolized by multiple CYP isoenzymes, leading to concerns about possible drug-drug interactions with other agents, such as the CYP3A substrate midazolam, which was increased by approximately 30% after 7 days of concomitant olorofim dosing.¹⁸¹

A phase 2b clinical trial (NCT03583164) is currently evaluating oral olorofim (up to 300 mg a day, guided by TDM) in patients with difficult-to-treat IFIs due to *L. prolificans*, *Scedosporium* spp, *Aspergillus* spp, and other resistant fungi. Several case reports presented in 2020 described successful treatment of infections due to *L. prolificans* and *Coccidioides* spp, with oral olorofim in individual patients participating in this trial.^{182–184}

Olorofim has been granted an orphan drug designation for coccidioidomycosis as well as a breakthrough drug designation for treatment of refractory/intolerant CNS coccidioidomycosis. In addition, it has an orphan drug designation for treatment of invasive aspergillosis and *Lomentospora/Scedosporium* infections. Olorofim also has been designated as a QIDP and breakthrough drug for treatment of difficult-to-treat mold infections, including invasive aspergillosis, scedosporiosis, lomentosporiosis, fusariosis, and infections due to *Scopulariopsis* spp.

Glycosylphosphatidylinositol Inhibitors

Fosmanogepix (APX001, Amplyx Pharmaceuticals, San Diego, CA) is a small molecule prodrug of the antifungal manogepix (APX001 A) that interferes with assembly of mannoproteins into cell wall glucan. By inhibiting the fungal Gwt1 enzyme, manogepix reduces cellular trafficking and anchoring of mannoproteins in cell membranes and the

outer cell wall, leading to a variety of detrimental effects on fungal cell integrity, growth, and virulence.^{44,185}

In vitro studies have shown that manogepix has potent activity against a variety of *Candida* spp, including *C auris* as well as *Cryptococcus neoformans*, *Aspergillus* spp (including azole-resistant and echinocandin-resistant organisms), and *L prolificans*, with variable activity against *Scedosporium* spp and *Fusarium* spp.^{44,45,186} Higher MICs generally have been observed for manogepix against Mucorales. Mutations in the Gwt1 target are likely to lead to manogepix resistance. This does not seem to impart cross-resistance with other antifungal classes. Some fluconazole-resistant *Candida* with efflux-related mutations, however, have exhibited higher MICs to manogepix; more data are needed to evaluate the clinical relevance of these findings.⁴⁴

Fosmanogepix is being studied in both IV and oral formulations. Data from humans suggests it has a half-life of 2 days to 2.5 days with linear pharmacokinetics and, in preclinical studies, appeared to be distributed well in tissues, including the eye and brain. It appears to be metabolized in the liver via several pathways and may be a substrate of several CYP isoenzymes, including CYP2C19, CYP2D6, CYP3A4, and CYP3A5.¹⁸⁷

Phase 2 clinical trials have focused on evaluating fosmanogepix for candidemia (NCT03604705), invasive *C auris* infections (NCT04148287), and invasive aspergillosis/rare mold infections (NCT04240886). In a small proof-of-concept phase 2, open-label study, 80% of non-neutropenic patients with candidemia experienced treatment success after 14 days of fosmanogepix (1 g twice daily IV for 1 day, followed by 600 mg daily IV or 700 mg daily orally after 3 days). AmB-resistant or anidulafungin-resistant *Candida* spp. infections occurred in 66% (N = 14) patients in the study; 71% (N=10) of these patients experienced treatment success. The most common treatment-emergent adverse effects were gastrointestinal disturbances.¹⁸⁸

Fosmanogepix has an orphan drug and QIDP/fast track FDA designation for the treatment of invasive candidiasis, invasive aspergillosis, coccidioidomycosis, cryptococcosis, and rare mold infections.¹⁸⁹

SUMMARY

This is an exciting time in antifungal development, with more than 7 agents in phase 2 or phase 3 clinical trials for a variety of IFIs. Polyenes, antimetabolites, azoles, and echinocandins all have a place in prevention or treatment of fungal infections. Given the challenges with dosing, pharmacokinetics, drug interactions, safety, and mycologic activity with each of these classes, however, there is room for new agents in the armamentarium. Additional studies are ongoing to optimize use of both new and older agents against highly morbid fungal infections.

CLINICS CARE POINTS

- Polyenes are the antifungal class with the broadest spectrum of activity and few drug-drug interactions but have poor oral bioavailability in currently approved forms. Side effects, such as nephrotoxicity, infusion-related reactions, and electrolyte disturbances, are a consideration when using agents in this class.
- Newer formulations of AmB include encochleated preparations, which may enable systemic concentrations with oral dosing.
- Azoles each have unique properties in regards to spectrum of activity, pharmacokinetics, drug-drug interactions, and safety profile. Availability in both oral and IV forms facilitates

administration of azoles in a variety of settings. TDM is recommended for several of the mold-active azoles to optimize efficacy and tolerability.

- Investigational azoles have greater selectivity for the target for lanosterol 14 α -demethylase and may have fewer off-target effects, leading to favorable safety profiles while retaining potent antifungal activity.
- Echinocandins target (1 \rightarrow 3)- β -D-glucan synthase in the fungal cell wall, leading to fungicidal activity against *Candida* spp and fungistatic activity against *Aspergillus* spp. They generally are well tolerated but are available only in IV formulations.
- A new echinocandin in development, rezafungin, has a much longer half-life than currently approved echinocandins, which will facilitate once-weekly dosing for IFIs.
- An investigational glucan synthase inhibitor, ibrexafungerp, is being studied in both IV and oral formulations and offers promise for candida infections, including *C auris*, as well as some invasive mold infections.
- Other agents include the antimetabolite 5-FC, which is used as adjunctive therapy in cryptococcal and candida infections. Its use is challenged by development of resistance and complexity of oral dosing due to its toxicity profile and need for TDM.
- Antifungals from novel classes, such as orotomides and glycosylphosphatidylinositol inhibitors, have entered phase 2 clinical trials and hopefully offer new alternatives with a wide spectrum of activity, IV and oral formulations, and favorable safety profiles.

DISCLOSURE

Dr M.D. Johnson has been a consultant to Cidara, Pfizer, and Astellas, and has research grants from Astellas, Scynexis, Charles River Laboratories, and Merck & Co.

REFERENCES

1. Warris A, Lehrnbecher T, Roilides E, et al. ESCMID-ECMM guideline: diagnosis and management of invasive aspergillosis in neonates and children. *Clin Microbiol Infect* 2019;25(9):1096–113.
2. Hope WW, Castagnola E, Groll AH, et al. ESCMID* guideline for the diagnosis and management of Candida diseases 2012: prevention and management of invasive infections in neonates and children caused by *Candida* spp. *Clin Microbiol Infect* 2012;18(Suppl 7):38–52.
3. Finkelstein A, Holz R. Aqueous pores created in thin lipid membranes by the polyene antibiotics nystatin and amphotericin B. *Membranes* 1973;2:377–408.
4. Nett JE, Andes DR. Antifungal agents: spectrum of activity, pharmacology, and clinical indications. *Infect Dis Clin North Am* 2016;30(1):51–83.
5. Chaabane F, Graf A, Jequier L, et al. Review on antifungal resistance mechanisms in the emerging pathogen *Candida auris*. *Front Microbiol* 2019;10:2788.
6. Seo K, Akiyoshi H, Ohnishi Y. Alteration of cell wall composition leads to amphotericin B resistance in *Aspergillus flavus*. *Microbiol Immunol* 1999;43(11):1017–25.
7. Nolte FS, Parkinson T, Falconer DJ, et al. Isolation and characterization of fluconazole- and amphotericin B-resistant *Candida albicans* from blood of two patients with leukemia. *Antimicrob Agents Chemother* 1997;41(1):196–9.
8. Escandón P, Chow NA, Caceres DH, et al. Molecular Epidemiology of *Candida auris* in Colombia reveals a highly related, countrywide colonization with regional patterns in amphotericin B resistance. *Clin Infect Dis* 2019;68(1):15–21.

9. Faustino C, Pinheiro L. Lipid systems for the delivery of amphotericin B in anti-fungal therapy. *Pharmaceutics* 2020;12(1):29.
10. Ashley ESD, Lewis R, Lewis JS, et al. Pharmacology of systemic antifungal agents. *Clin Infect Dis* 2006;43(Supplement_1):S28–39.
11. Wu J-Q, Shao K, Wang X, et al. In vitro and in vivo evidence for amphotericin B as a P-glycoprotein substrate on the blood-brain barrier. *Antimicrob Agents Chemother* 2014;58(8):4464–9.
12. Bekersky I, Fielding RM, Dressler DE, et al. Pharmacokinetics, excretion, and mass balance of liposomal amphotericin B (AmBisome) and amphotericin B deoxycholate in humans. *Antimicrob Agents Chemother* 2002;46(3):828–33.
13. Stevens DA, Clemons KV, Martinez M, et al. The brain, amphotericin B, and P-glycoprotein. *Antimicrob Agents Chemother* 2015;59:1386.
14. Al-Nakeeb Z, Petraitis V, Goodwin J, et al. Pharmacodynamics of amphotericin B deoxycholate, amphotericin B lipid complex, and liposomal amphotericin B against *Aspergillus fumigatus*. *Antimicrob Agents Chemother* 2015;59(5):2735–45.
15. Adedoyin A, Swenson CE, Bolcsak LE, et al. A pharmacokinetic study of amphotericin B lipid complex injection (Abelcet) in patients with definite or probable systemic fungal infections. *Antimicrob Agents Chemother* 2000;44(10):2900–2.
16. Skipper CP, Atukunda M, Stadelman A, et al. Phase I EnACT trial of the safety and tolerability of a novel oral formulation of amphotericin B. *Antimicrob Agents Chemother* 2020;64(10):e00838.
17. Vermes A, Guchelaar HJ, Dankert J. Flucytosine: a review of its pharmacology, clinical indications, pharmacokinetics, toxicity and drug interactions. *J Antimicrob Chemother* 2000;46(2):171–9.
18. Felton T, Troke PF, Hope WW. Tissue penetration of antifungal agents. *Clin Microbiol Rev* 2014;27(1):68–88.
19. Sinnolareddy MG, Roberts JA, Lipman J, et al. Pharmacokinetic variability and exposures of fluconazole, anidulafungin, and caspofungin in intensive care unit patients: Data from multinational Defining Antibiotic Levels in Intensive care unit (DALI) patients Study. *Crit Care* 2015;19:33.
20. Andes D, Kovanda L, Desai A, et al. Isavuconazole concentration in real-world practice: consistency with results from clinical trials. *Antimicrob Agents Chemother* 2018;62(7):e00585.
21. Yamazaki T, Desai A, Goldwater R, et al. Pharmacokinetic interactions between isavuconazole and the drug transporter substrates atorvastatin, digoxin, metformin, and methotrexate in healthy subjects. *Clin Pharmacol Drug Dev* 2017;6(1):66–75.
22. Desai A, Kovanda L, Kowalski D, et al. Population Pharmacokinetics of Isavuconazole from Phase 1 and Phase 3 (SECURE) trials in adults and target attainment in patients with invasive infections due to *aspergillus* and other filamentous fungi. *Antimicrob Agents Chemother* 2016;60(9):5483–91.
23. Lestner J, Hope WW. Itraconazole: an update on pharmacology and clinical use for treatment of invasive and allergic fungal infections. *Expert Opin Drug Metab Toxicol* 2013;9(7):911–26.
24. Daneshmend TK, Warnock DW, Turner A, et al. Pharmacokinetics of ketoconazole in normal subjects. *J Antimicrob Chemother* 1981;8(4):299–304.
25. Van Tyle JH. Ketoconazole. Mechanism of action, spectrum of activity, pharmacokinetics, drug interactions, adverse reactions and therapeutic use. *Pharmacotherapy* 1984;4(6):343–73.

26. Kaeser B, Zandt H, Bour F, et al. Drug-drug interaction study of ketoconazole and ritonavir-boosted saquinavir. *Antimicrob Agents Chemother* 2009;53(2):609–14.
27. Chen L, Krekels EHJ, Verweij PE, et al. Pharmacokinetics and Pharmacodynamics of Posaconazole. *Drugs* 2020;80(7):671–95.
28. Purkins L, Wood N, Kleinermans D, et al. Voriconazole does not affect the steady-state pharmacokinetics of digoxin. *Br J Clin Pharmacol* 2003;56(Suppl 1):45–50.
29. Brand SR, Degenhardt TP, Person K, et al. A phase 2, randomized, double-blind, placebo-controlled, dose-ranging study to evaluate the efficacy and safety of orally administered VT-1161 in the treatment of recurrent vulvovaginal candidiasis. *Am J Obstet Gynecol* 2018;218(6):624.e1–9.
30. Warrilow AGS, Hull CM, Parker JE, et al. The clinical candidate VT-1161 is a highly potent inhibitor of *Candida albicans* CYP51 but fails to bind the human enzyme. *Antimicrob Agents Chemother* 2014;58(12):7121–7.
31. Shubitz LF, Roy ME, Trinh HT, et al. Efficacy of the Investigational Antifungal VT-1161 in Treating Naturally Occurring Coccidioidomycosis in Dogs. *Antimicrob Agents Chemother* 2017;61(5):e00111.
32. Garvey EP, Hoekstra WJ, Schotzinger RJ, et al. Efficacy of the clinical agent VT-1161 against fluconazole-sensitive and -resistant *Candida albicans* in a murine model of vaginal candidiasis. *Antimicrob Agents Chemother* 2015;59(9):5567–73.
33. Kofla G, Ruhnke M. Pharmacology and metabolism of anidulafungin, caspofungin and micafungin in the treatment of invasive candidosis: review of the literature. *Eur J Med Res* 2011;16(4):159–66.
34. Stone JA, Xu X, Winchell GA, et al. Disposition of caspofungin: role of distribution in determining pharmacokinetics in plasma. *Antimicrob Agents Chemother* 2004;48(3):815–23.
35. Sandison T, Ong V, Lee J, et al. Safety and pharmacokinetics of CD101 IV, a novel echinocandin, in healthy adults. *Antimicrob Agents Chemother* 2017;61(2):e01627.
36. Thompson GR, Soriano A, Skoutelis A, et al. Rezafungin versus Caspofungin in a Phase 2, randomized, double-blind study for the treatment of Candidemia and Invasive Candidiasis- The STRIVE Trial. *Clin Infect Dis* 2020. <https://doi.org/10.1093/cid/ciaa1380>.
37. Ong V, James KD, Smith S, et al. Pharmacokinetics of the novel echinocandin CD101 in multiple animal species. *Antimicrob Agents Chemother* 2017;61(4):e01626.
38. Ong V, Flanagan S, Sandison T, et al. 1286. Pharmacokinetics, excretion, and mass balance of [14C]-Rezafungin Following Intravenous (IV) Administration in Healthy Adults. *Open Forum Infect Dis* 2020;7(Suppl 1):S658.
39. Azie N, Angulo D, Dehn B, et al. Oral Ibrexafungerp: an investigational agent for the treatment of vulvovaginal candidiasis. *Expert Opin Investig Drugs* 2020;29(9):893–900.
40. Wring SA, Randolph R, Park S, et al. Preclinical pharmacokinetics and pharmacodynamic target of SCY-078, a first-in-class orally active antifungal glucan synthesis inhibitor, in murine models of disseminated candidiasis. *Antimicrob Agents Chemother* 2017;61(4):e02068.
41. Wring S, Murphy G, Attee G, et al. Clinical pharmacokinetics and drug-drug interaction potential for Coadministered SCY-078, an oral fungicidal glucan synthase inhibitor, and tacrolimus. *Clin Pharmacol Drug Dev* 2019;8(1):60–9.

42. Wiederhold NP. Review of the novel investigational antifungal Olorofim. *J Fungi (Basel)* 2020;6(3):122.
43. Kennedy T, Allen G, Steiner J, et al. Multiple dose pharmacokinetics of an immediate-release tablet formulation of F901318 in healthy male and female subjects. In: Proceedings of the 27th European congress of clinical microbiology and infectious Diseases. 2017.
44. Shaw KJ, Ibrahim AS. Fosmanogepix: a review of the first-in-class broad spectrum agent for the treatment of invasive fungal infections. *J Fungi (Basel)* 2020; 6(4):239.
45. Zhao M, Lepak AJ, Marchillo K, et al. APX001 Pharmacokinetic/Pharmacodynamic target determination against *Aspergillus fumigatus* in an in vivo model of invasive pulmonary aspergillosis. *Antimicrob Agents Chemother* 2019. <https://doi.org/10.1128/aac.02372-18>.
46. Hodges MR, Ople E, Shaw KJ, et al. Phase 1 study to assess safety, tolerability and pharmacokinetics of single and multiple oral doses of APX001 and to investigate the effect of food on APX001 bioavailability. *Open Forum Infect Dis* 2017; 4(Suppl 1):S534.
47. Gómez-López A. Antifungal therapeutic drug monitoring: focus on drugs without a clear recommendation. *Clin Microbiol Infect* 2020;26(11):1481–7.
48. Lepak AJ, Andes DR. Antifungal pharmacokinetics and pharmacodynamics. *Cold Spring Harb Perspect Med* 2014;5(5):a019653.
49. Hope WW, McEntee L, Livermore J, et al. Pharmacodynamics of the orotomides against *Aspergillus fumigatus*: new opportunities for treatment of multidrug-resistant fungal disease. *MBio* 2017;8(4):e01157.
50. Duckwall MJ, Gales MA, Gales BJ. Inhaled Amphotericin B as aspergillosis prophylaxis in hematologic disease: an update. *Microbiol Insights* 2019;12: 1178636119869937.
51. Venanzi E, Martín-Dávila P, López J, et al. aerosolized lipid amphotericin B for complementary therapy and/or secondary prophylaxis in patients with invasive pulmonary aspergillosis: a single-center experience. *Mycopathologia* 2019; 184(2):239–50.
52. Safdar A, Rodriguez GH. Aerosolized amphotericin B lipid complex as adjunctive treatment for fungal lung infection in patients with cancer-related immunosuppression and recipients of hematopoietic stem cell transplantation. *Pharmacotherapy* 2013;33(10):1035–43.
53. Mohr J, Johnson M, Cooper T, et al. Current options in antifungal pharmacotherapy. *Pharmacotherapy* 2008;28(5):614–45.
54. Hamill RJ. Amphotericin B formulations: a comparative review of efficacy and toxicity. *Drugs* 2013;73(9):919–34.
55. Chai LYA, Netea MG, Tai BC, et al. An elevated pro-inflammatory cytokine response is linked to development of amphotericin B-induced nephrotoxicity. *J Antimicrob Chemother* 2013;68(7):1655–9.
56. Botero Aguirre JP, Restrepo Hamid AM. Amphotericin B deoxycholate versus liposomal amphotericin B: effects on kidney function. *Cochrane Database Syst Rev* 2015;(11):CD010481.
57. Steimbach LM, Tonin FS, Virtuoso S, et al. Efficacy and safety of amphotericin B lipid-based formulations-A systematic review and meta-analysis. *Mycoses* 2017;60(3):146–54.
58. Ellis D. Amphotericin B: spectrum and resistance. *J Antimicrob Chemother* 2002;49(Suppl 1):7–10.

59. Saravolatz LD, Ostrosky-Zeichner L, Marr KA, et al. Amphotericin B: time for a new "Gold Standard". *Clin Infect Dis* 2003;37(3):415–25.
60. Tortorano AM, Richardson M, Roilides E, et al. ESCMID and ECMM joint guidelines on diagnosis and management of hyalohyphomycosis: *Fusarium* spp., *Scedosporium* spp. and others. *Clin Microbiol Infect* 2014;20(Suppl 3):27–46.
61. Ullmann AJ, Aguado JM, Arikan-Akdagli S, et al. Diagnosis and management of *Aspergillus* diseases: executive summary of the 2017 ESCMID-ECMM-ERS guideline. *Clin Microbiol Infect* 2018;24(Suppl 1):e1–38.
62. Cuddihy G, Wasan EK, Di Y, et al. The development of oral amphotericin b to treat systemic fungal and parasitic infections: has the myth been finally realized? *Pharmaceutics* 2019;11(3):99.
63. Aigner M, Lass-Flörl C. Encocleated Amphotericin B: Is the oral availability of amphotericin b finally reached? *J Fungi (Basel)* 2020;6(2):66.
64. Zarif L, Graybill JR, Perlin D, et al. Antifungal activity of amphotericin B cochleates against *Candida albicans* infection in a mouse model. *Antimicrob Agents Chemother* 2000;44(6):1463–9.
65. Segarra I, Movshin DA, Zarif L. Pharmacokinetics and tissue distribution after intravenous administration of a single dose of amphotericin B cochleates, a new lipid-based delivery system. *J Pharm Sci* 2002;91(8):1827–37.
66. Lu R, Hollingsworth C, Qiu J, et al. Efficacy of oral encocleated amphotericin b in a mouse model of cryptococcal meningoencephalitis. *MBio* 2019;10(3):e00724.
67. NCT02971007. Safety and efficacy of oral encocleated amphotericin B (CAMB/MAT2203) in the treatment of vulvovaginal Candidiasis. U.S. National Library of Medicine ClinicalTrials.gov. Available at: <https://clinicaltrials.gov/ct2/show/NCT02971007>. Accessed February 15, 2021.
68. Dixon DM, Polak A. In vitro and in vivo drug studies with three agents of central nervous system phaeohyphomycosis. *Chemotherapy* 1987;33(2):129–40.
69. Pfaller MA, Messer SA, Boyken L, et al. In vitro activities of 5-fluorocytosine against 8,803 clinical isolates of *Candida* spp.: global assessment of primary resistance using National Committee for Clinical Laboratory Standards susceptibility testing methods. *Antimicrob Agents Chemother* 2002;46(11):3518–21.
70. Stamm AM, Diasio RB, Dismukes WE, et al. Toxicity of amphotericin B plus flucytosine in 194 patients with cryptococcal meningitis. *Am J Med* 1987;83(2):236–42.
71. Perfect JR, Dismukes WE, Dromer F, et al. Clinical practice guidelines for the management of cryptococcal disease: 2010 update by the Infectious Diseases Society of America. *Clin Infect Dis* 2010;50(3):291–322.
72. Pappas PG, Kauffman CA, Andes DR, et al. Clinical practice guideline for the management of candidiasis: 2016 update by the infectious diseases society of America. *Clin Infect Dis* 2016;62(4):e1–50.
73. Queiroz-Telles F, de Hoog S, Santos DWCL, et al. Chromoblastomycosis. *Clin Microbiol Rev* 2017;30(1):233–76.
74. Chowdhary A, Meis JF, Guarro J, et al. ESCMID and ECMM joint clinical guidelines for the diagnosis and management of systemic phaeohyphomycosis: diseases caused by black fungi. *Clin Microbiol Infect* 2014;20(Suppl 3):47–75.
75. Leonardelli F, Macedo D, Dudiuk C, et al. *Aspergillus fumigatus* intrinsic fluconazole resistance is due to the naturally occurring T301I substitution in Cyp51Ap. *Antimicrob Agents Chemother* 2016;60(9):5420–6.

76. Beardsley J, Halliday CL, Chen SC-A, et al. Responding to the emergence of antifungal drug resistance: perspectives from the bench and the bedside. *Future Microbiol* 2018;13:1175–91.
77. Pfaller MA, Diekema DJ, Turnidge JD, et al. Twenty years of the sentry antifungal surveillance program: results for *Candida* species from 1997–2016. *Open Forum Infect Dis* 2019;6(Suppl 1):S79–94.
78. Snelders E, Huis In 't Veld RAG, Rijs AJMM, et al. Possible environmental origin of resistance of *Aspergillus fumigatus* to medical triazoles. *Appl Environ Microbiol* 2009;75(12):4053–7.
79. Ricardo E, Grenouillet F, Miranda IM, et al. Mechanisms of acquired *in vivo* and *in vitro* resistance to voriconazole by *Candida Krusei* following exposure to sub-optimal drug concentration. *Antimicrob Agents Chemother* 2020;64(4):e01651.
80. Flowers SA, Colón B, Whaley SG, et al. Contribution of clinically derived mutations in ERG11 to azole resistance in *Candida albicans*. *Antimicrob Agents Chemother* 2015;59(1):450–60.
81. Resendiz-Sharpe A, Mercier T, Lestrade PPA, et al. Prevalence of voriconazole-resistant invasive aspergillosis and its impact on mortality in haematology patients. *J Antimicrob Chemother* 2019;74(9):2759–66.
82. Lestrade PP, Bentvelsen RG, Schauvlyliege AFAD, et al. Voriconazole resistance and mortality in invasive aspergillosis: a multicenter retrospective cohort study. *Clin Infect Dis* 2019;68(9):1463–71.
83. Andes D, Pascual A, Marchetti O. Antifungal therapeutic drug monitoring: established and emerging indications. *Antimicrob Agents Chemother* 2009;53(1):24–34.
84. Zurl C, Waller M, Schwameis F, et al. Isavuconazole treatment in a mixed patient cohort with invasive fungal infections: outcome, tolerability and clinical implications of Isavuconazole plasma concentrations. *J Fungi (Basel)* 2020;6(2):90.
85. Rouzaud C, Jullien V, Herbrecht A, et al. Isavuconazole diffusion in infected human brain. *Antimicrob Agents Chemother* 2019;63(10):e02474.
86. Kosmidis C, Otu A, Moore CB, et al. Isavuconazole therapeutic drug monitoring during long-term treatment for chronic pulmonary aspergillosis. *Antimicrob Agents Chemother* 2020;65(1):e01511.
87. Abuhelwa AY, Foster DJR, Mudge S, et al. Population pharmacokinetic modeling of itraconazole and hydroxyitraconazole for oral SUBA-itraconazole and sporanox capsule formulations in healthy subjects in fed and fasted states. *Antimicrob Agents Chemother* 2015;59(9):5681–96.
88. Duflot T, Schrapp A, Bellien J, et al. Impact of CYP3A4 genotype on voriconazole exposure. *Clin Pharmacol Ther* 2018;103(2):185–6.
89. Moriyama B, Obeng AO, Barbarino J, et al. Clinical Pharmacogenetics Implementation Consortium (CPIC) guidelines for CYP2C19 and voriconazole therapy. *Clin Pharmacol Ther* 2017;102(1):45–51.
90. Pfizer Inc. Vfend (voriconazole) [package insert]. New York, NY: Pfizer, Inc; 2019.
91. Owusu Obeng A, Egelund EF, Alsultan A, et al. CYP2C19 polymorphisms and therapeutic drug monitoring of voriconazole: are we ready for clinical implementation of pharmacogenomics? *Pharmacotherapy* 2014;34(7):703–18.
92. Spiert I, Annaert P, Meersseman P, et al. Pharmacokinetics of caspofungin and voriconazole in critically ill patients during extracorporeal membrane oxygenation. *J Antimicrob Chemother* 2009;63(4):767–70.

93. Davies-Vorbrodt S, Ito JI, Tegtmeier BR, et al. Voriconazole serum concentrations in obese and overweight immunocompromised patients: a retrospective review. *Pharmacotherapy* 2013;33(1):22–30.
94. Kosalke E, Kraft S, Smith J, et al. Evaluation of the effect of obesity on voriconazole serum concentrations. *J Antimicrob Chemother* 2012;67(12):2957–62.
95. Courtney R, Pai S, Laughlin M, et al. Pharmacokinetics, safety, and tolerability of oral posaconazole administered in single and multiple doses in healthy adults. *Antimicrob Agents Chemother* 2003;47(9):2788–95.
96. Krishna G, Moton A, Ma L, et al. Pharmacokinetics and absorption of posaconazole oral suspension under various gastric conditions in healthy volunteers. *Antimicrob Agents Chemother* 2009;53(3):958–66.
97. Wiederhold NP. Pharmacokinetics and safety of posaconazole delayed-release tablets for invasive fungal infections. *Clin Pharmacol* 2016;8:1–8.
98. Rybak JM, Marx KR, Nishimoto AT, et al. Isavuconazole: pharmacology, pharmacodynamics, and current clinical experience with a New Triazole antifungal agent. *Pharmacotherapy* 2015;35(11):1037–51.
99. Brüggemann RJM, Alffenaar J-WC, Blijlevens NMA, et al. Clinical relevance of the pharmacokinetic interactions of azole antifungal drugs with other coadministered agents. *Clin Infect Dis* 2009;48(10):1441–58.
100. Andes D, Azie N, Yang H, et al. Drug-drug interaction associated with mold-active Triazoles among hospitalized patients. *Antimicrob Agents Chemother* 2016;60(6):3398–406.
101. Stemler J, Koehler P, Maurer C, et al. Antifungal prophylaxis and novel drugs in acute myeloid leukemia: the midostaurin and posaconazole dilemma. *Ann Hematol* 2020;99(7):1429–40.
102. Mellinghoff SC, Bassetti M, Dörfel D, et al. Isavuconazole shortens the QTc interval. *Mycoses* 2018;61(4):256–60.
103. Benitez LL, Carver PL. Adverse effects associated with long-term administration of azole antifungal agents. *Drugs* 2019;79(8):833–53.
104. Bérard A, Sheehy O, Zhao J-P, et al. Associations between low- and high-dose oral fluconazole and pregnancy outcomes: 3 nested case-control studies. *CMAJ* 2019;191(7):E179–87.
105. Mølgaard-Nielsen D, Svanström H, Melbye M, et al. Association between use of oral fluconazole during pregnancy and risk of Spontaneous Abortion and Stillbirth. *JAMA* 2016;315(1):58–67.
106. Budani MC, Fensore S, Di Marzio M, et al. Maternal use of fluconazole and congenital malformations in the progeny: a meta-analysis of the literature. *Reprod Toxicol* 2020;100:42–51.
107. Liu D, Zhang C, Wu L, et al. Fetal outcomes after maternal exposure to oral anti-fungal agents during pregnancy: a systematic review and meta-analysis. *Int J Gynaecol Obstet* 2020;148(1):6–13.
108. Pascual A, Calandra T, Bolay S, et al. Voriconazole therapeutic drug monitoring in patients with invasive mycoses improves efficacy and safety outcomes. *Clin Infect Dis* 2008;46(2):201–11.
109. Jin H, Wang T, Falcione BA, et al. Trough concentration of voriconazole and its relationship with efficacy and safety: a systematic review and meta-analysis. *J Antimicrob Chemother* 2016;71(7):1772–85.
110. Tang H, Shi W, Song Y, et al. Voriconazole exposure and risk of cutaneous squamous cell carcinoma among lung or hematopoietic cell transplant patients: a systematic review and meta-analysis. *J Am Acad Dermatol* 2019;80(2):500–7.e10.

111. Nguyen M-VH, Davis MR, Wittenberg R, et al. posaconazole serum drug levels associated with pseudohyperaldosteronism. *Clin Infect Dis* 2020;70(12):2593–8.
112. Beck KR, Telisman L, van Koppen CJ, et al. Molecular mechanisms of posaconazole- and itraconazole-induced pseudohyperaldosteronism and assessment of other systemically used azole antifungals. *J Steroid Biochem Mol Biol* 2020; 199:105605.
113. Teaford HR, Abu Saleh OM, Villarraga HR, et al. The many faces of itraconazole cardiac toxicity. *Mayo Clin Proc Innov Qual Outcomes* 2020;4(5):588–94.
114. Hauben M, Hung EY. A quantitative analysis of the spontaneous reporting of congestive heart failure-related adverse events with systemic anti-fungal drugs. *J Clin Pharmacol* 2013;53(7):762–72.
115. Kiser TH, Fish DN, Aquilante CL, et al. Evaluation of sulfobutylether- β -cyclodextrin (SBECD) accumulation and voriconazole pharmacokinetics in critically ill patients undergoing continuous renal replacement therapy. *Crit Care* 2015;19:32.
116. Kim S-H, Kwon J-C, Park C, et al. Therapeutic drug monitoring and safety of intravenous voriconazole formulated with sulfobutylether β -cyclodextrin in haematological patients with renal impairment. *Mycoses* 2016;59(10):644–51.
117. Luke DR, Tomaszewski K, Damle B, et al. Review of the basic and clinical pharmacology of sulfobutylether-beta-cyclodextrin (SBECD). *J Pharm Sci* 2010; 99(8):3291–301.
118. Neofytos D, Lombardi LR, Shields RK, et al. Administration of voriconazole in patients with renal dysfunction. *Clin Infect Dis* 2012;54(7):913–21.
119. Turner RB, Martello JL, Malhotra A. Worsening renal function in patients with baseline renal impairment treated with intravenous voriconazole: a systematic review. *Int J Antimicrob Agents* 2015;46(4):362–6.
120. Stella VJ, He Q. Cyclodextrins. *Toxicol Pathol* 2008;36(1):30–42.
121. Stevens DA. Itraconazole in cyclodextrin solution. *Pharmacotherapy* 1999;19(5): 603–11.
122. Vallabhaneni S, Baggs J, Tsay S, et al. Trends in antifungal use in US hospitals, 2006–12. *J Antimicrob Chemother* 2018;73(10):2867–75.
123. Chapman SW, Dismukes WE, Proia LA, et al. Clinical practice guidelines for the management of blastomycosis: 2008 update by the infectious diseases society of America. *Clin Infect Dis* 2008;46(12):1801–12.
124. Galgiani JN, Ampel NM, Blair JE, et al. 2016 Infectious Diseases Society of America (IDSA) clinical practice guideline for the treatment of coccidioidomycosis. *Clin Infect Dis* 2016;63(6):e112–46.
125. Wheat LJ, Freifeld AG, Kleiman MB, et al. Clinical practice guidelines for the management of patients with histoplasmosis: 2007 update by the Infectious Diseases Society of America. *Clin Infect Dis* 2007;45(7):807–25.
126. Kauffman CA, Bustamante B, Chapman SW, et al. Clinical practice guidelines for the management of sporotrichosis: 2007 update by the Infectious Diseases Society of America. *Clin Infect Dis* 2007;45(10):1255–65.
127. Patterson TF, Thompson GR 3rd, Denning DW, et al. Practice guidelines for the diagnosis and management of aspergillosis: 2016 update by the infectious diseases society of America. *Clin Infect Dis* 2016;63(4):e1–60.
128. Cornely OA, Alastruey-Izquierdo A, Arenz D, et al. Global guideline for the diagnosis and management of mucormycosis: an initiative of the European Confederation of Medical Mycology in cooperation with the Mycoses Study Group Education and Research Consortium. *Lancet Infect Dis* 2019;19(12):e405–21.

129. Ullmann AJ, Lipton JH, Vesole DH, et al. Posaconazole or fluconazole for prophylaxis in severe graft-versus-host disease. *N Engl J Med* 2007;356(4):335–47.
130. Cornely OA, Maertens J, Winston DJ, et al. Posaconazole vs. fluconazole or itraconazole prophylaxis in patients with neutropenia. *N Engl J Med* 2007;356(4):348–59.
131. Proia LA, Harnisch DO. Successful use of posaconazole for treatment of blastomycosis. *Antimicrob Agents Chemother* 2012;56(7):4029.
132. Heidari A, Quinlan M, Benjamin DJ, et al. Isavuconazole in the treatment of coccidioidal meningitis. *Antimicrob Agents Chemother* 2019;63(3):e02232.
133. Yates CM, Garvey EP, Shaver SR, et al. Design and optimization of highly-selective, broad spectrum fungal CYP51 inhibitors. *Bioorg Med Chem Lett* 2017;27(15):3243–8.
134. Schell WA, Jones AM, Garvey EP, et al. Fungal CYP51 inhibitors VT-1161 and VT-1129 exhibit strong in vitro activity against *Candida Glabrata* and *C. Krusei* isolates clinically resistant to azole and echinocandin antifungal compounds. *Antimicrob Agents Chemother* 2017;61(3):e01817.
135. Wiederhold NP. The antifungal arsenal: alternative drugs and future targets. *Int J Antimicrob Agents* 2018;51(3):333–9.
136. Wiederhold NP, Lockhart SR, Najvar LK, et al. The fungal Cyp51-specific inhibitor VT-1598 demonstrates in vitro and in vivo activity against *Candida auris*. *Antimicrob Agents Chemother* 2019;63(3):e02233.
137. Wiederhold NP, Patterson HP, Tran BH, et al. Fungal-specific Cyp51 inhibitor VT-1598 demonstrates in vitro activity against *Candida* and *Cryptococcus* species, endemic fungi, including *Coccidioides* species, *Aspergillus* species and *Rhizopus arrhizus*. *J Antimicrob Chemother* 2018;73(2):404–8.
138. Wiederhold NP, Xu X, Wang A, et al. In vivo efficacy of VT-1129 against experimental cryptococcal meningitis with the use of a loading dose-maintenance dose administration strategy. *Antimicrob Agents Chemother* 2018;62(11):e01315.
139. Jiménez-Ortigosa C, Perez WB, Angulo D, et al. De novo acquisition of resistance to SCY-078 in *Candida glabrata* involves FKS mutations that both overlap and are distinct from those conferring echinocandin resistance. *Antimicrob Agents Chemother* 2017;61(9):e00833.
140. Espinel-Ingroff A, Cantón E, Pemán J. Antifungal resistance among less prevalent candida non-albicans and other yeasts versus established and under development agents: a literature review. *J Fungi (Basel)* 2021;7(1):24.
141. Eschenauer G, Depestel DD, Carver PL. Comparison of echinocandin antifungals. *Ther Clin Risk Manag* 2007;3(1):71–97.
142. Alexander BD, Johnson MD, Pfeiffer CD, et al. Increasing echinocandin resistance in *Candida glabrata*: clinical failure correlates with presence of FKS mutations and elevated minimum inhibitory concentrations. *Clin Infect Dis* 2013;56(12):1724–32.
143. Shields RK, Nguyen MH, Press EG, et al. The presence of an FKS mutation rather than MIC is an independent risk factor for failure of echinocandin therapy among patients with invasive candidiasis due to *Candida glabrata*. *Antimicrob Agents Chemother* 2012;56(9):4862–9.
144. Arendrup MC, Dzajic E, Jensen RH, et al. Epidemiological changes with potential implication for antifungal prescription recommendations for fungaemia: data from a nationwide fungaemia surveillance programme. *Clin Microbiol Infect* 2013;19(8):E343–53.

145. Vossen MG, Knafl D, Haidinger M, et al. Micafungin plasma levels are not affected by continuous renal replacement therapy: experience in critically ill patients. *Antimicrob Agents Chemother* 2017;61(8):e02425.
146. Honore PM, De Bels D, Gutierrez LB, et al. Optimizing micafungin dosing in critically ill patients: what about extracorporeal therapies? *Crit Care* 2018;22:289.
147. Autmizguine J, Hornik CP, Benjamin DK Jr, et al. Pharmacokinetics and safety of micafungin in infants supported with extracorporeal membrane oxygenation. *Pediatr Infect Dis J* 2016;35(11):1204–10.
148. Wang Q, Zhang Z, Liu D, et al. Population pharmacokinetics of caspofungin among extracorporeal membrane oxygenation patients during the postoperative period of lung transplantation. *Antimicrob Agents Chemother* 2020;64(11): e00687.
149. Aguilar G, Ferriols R, Carbonell JA, et al. Pharmacokinetics of anidulafungin during venovenous extracorporeal membrane oxygenation. *Crit Care* 2016; 20(1):325.
150. Watt KM, Cohen-Wolkowicz M, Williams DC, et al. Antifungal Extraction by the Extracorporeal Membrane Oxygenation Circuit. *J Extra Corpor Technol* 2017; 49(3):150–9.
151. Wasmann RE, Smit C, Ter Heine R, et al. Pharmacokinetics and probability of target attainment for micafungin in normal-weight and morbidly obese adults. *J Antimicrob Chemother* 2019;74(4):978–85.
152. Wasmann RE, Ter Heine R, van Dongen EP, et al. Pharmacokinetics of anidulafungin in obese and normal-weight adults. *Antimicrob Agents Chemother* 2018; 62(7):e00063.
153. Hall RG 2nd, Swancutt MA, Meek C, et al. Weight drives caspofungin pharmacokinetic variability in overweight and obese people: fractal power signatures beyond two-thirds or three-fourths. *Antimicrob Agents Chemother* 2013;57(5): 2259–64.
154. Märtsönen A-G, van der Elst KCM, Veringa A, et al. Caspofungin weight-based dosing supported by a population pharmacokinetic model in critically ill patients. *Antimicrob Agents Chemother* 2020;64(9):e00905.
155. Gustot T, Ter Heine R, Brauns E, et al. Caspofungin dosage adjustments are not required for patients with Child-Pugh B or C cirrhosis. *J Antimicrob Chemother* 2018;73(9):2493–6.
156. Pappas PG, Rotstein CMF, Betts RF, et al. Micafungin versus caspofungin for treatment of candidemia and other forms of invasive candidiasis. *Clin Infect Dis* 2007;45(7):883–93.
157. Reboli AC, Rotstein C, Pappas PG, et al. Anidulafungin versus fluconazole for invasive candidiasis. *N Engl J Med* 2007;356(24):2472–82.
158. Mora-Duarte J, Betts R, Rotstein C, et al. Comparison of caspofungin and amphotericin B for invasive candidiasis. *N Engl J Med* 2002;347(25):2020–9.
159. Bormann AM, Morrison VA. Review of the pharmacology and clinical studies of micafungin. *Drug Des Devel Ther* 2009;3:295–302.
160. Schneeweiss S, Carver PL, Datta K, et al. Long-term risk of hepatocellular carcinoma mortality in 23220 hospitalized patients treated with micafungin or other parenteral antifungals. *J Antimicrob Chemother* 2020;75(1):221–8.
161. Schneeweiss S, Carver PL, Datta K, et al. Short-term risk of liver and renal injury in hospitalized patients using micafungin: a multicentre cohort study. *J Antimicrob Chemother* 2016;71(10):2938–44.
162. van Burik J-AH, Ratanatharathorn V, Stepan DE, et al. Micafungin versus fluconazole for prophylaxis against invasive fungal infections during neutropenia in

- patients undergoing hematopoietic stem cell transplantation. *Clin Infect Dis* 2004;39(10):1407–16.
163. Walsh TJ, Teppler H, Donowitz GR, et al. Caspofungin versus liposomal amphotericin B for empirical antifungal therapy in patients with persistent fever and neutropenia. *N Engl J Med* 2004;351(14):1391–402.
 164. Flanagan S, Goodman DB, Jandourek A, et al. Lack of effect of rezafungin on QT/QTc interval in healthy subjects. *Clin Pharmacol Drug Dev* 2020;9(4):456–65.
 165. Nyirjesy P, Alessio C, Jandourek A, et al. CD101 topical compared with oral fluconazole for acute vulvovaginal candidiasis: a randomized controlled trial. *J Low Genit Tract Dis* 2019;23(3):226–9.
 166. Ham YY, Lewis JS, Thompson GR. Rezafungin: a novel antifungal for the treatment of invasive candidiasis. *Future Microbiol* 2021;16:27–36.
 167. Pfaller MA, Messer SA, Rhomberg PR, et al. Differential activity of the oral glucan synthase inhibitor SCY-078 against wild-type and echinocandin-resistant strains of *Candida* Species. *Antimicrob Agents Chemother* 2017;61(8):e00161.
 168. Arendrup MC, Jørgensen KM, Hare RK, et al. In vitro activity of Ibrexafungerp (SCY-078) against *Candida auris* Isolates as Determined by EUCAST methodology and comparison with activity against *C. albicans* and *C. glabrata* and with the activities of six comparator agents. *Antimicrob Agents Chemother* 2020;64(3):e02136.
 169. Lamothe F, Alexander BD. Antifungal activities of SCY-078 (MK-3118) and standard antifungal agents against clinical non-Aspergillus mold isolates. *Antimicrob Agents Chemother* 2015;59(7):4308–11.
 170. SCYNEXIS Announces Advancement of Ibrexafungerp's intravenous formulation to clinical stage and provides further updates on its clinical studies in patients with life-threatening fungal infections : SCYNEXIS, Inc. (SCYX). Available at: <https://www.scynexis.com/news-media/press-releases/detail/226/scynexis-announces-advancement-of-ibrexafungerps>. Accessed February 19, 2021.
 171. Wring S, Borroto-Esoda K, Solon E, et al. SCY-078, a novel fungicidal agent, demonstrates distribution to tissues associated with fungal infections during mass balance studies with intravenous and oral [¹⁴C]SCY-078 in Albino and Pigmented Rats. *Antimicrob Agents Chemother* 2019;63(2):e02119.
 172. Lee A, Prideaux B, Zimmerman M, et al. Penetration of Ibrexafungerp (Formerly SCY-078) at the Site of Infection in an Intra-abdominal Candidiasis Mouse Model. *Antimicrob Agents Chemother* 2020;64(3):e02268.
 173. Wring S, Murphy G, Attee G, et al. Lack of Impact by SCY-078, a first-in-class oral fungicidal glucan synthase inhibitor, on the pharmacokinetics of rosiglitazone, a substrate for CYP450 2C8, supports the low risk for clinically relevant metabolic drug-drug interactions. *J Clin Pharmacol* 2018;58(10):1305–13.
 174. Spec A, Pullman J, Thompson GR, et al. MSG-10: a Phase 2 study of oral ibrexafungerp (SCY-078) following initial echinocandin therapy in non-neutropenic patients with invasive candidiasis. *J Antimicrob Chemother* 2019;74(10):3056–62.
 175. Alexander BD, Cornely OA, Pappag PG, et al. Efficacy and safety of oral ibrexafungerp in 41 patients with refractory fungal diseases, interim analysis of a phase 3 open-label study (FURI). In: Proceedings of IDWeek2020. 2020.
 176. Van Daele R, Spriet I, Wauters J, et al. Antifungal drugs: what brings the future? *Med Mycol* 2019;57(Supplement_3):S328–43.
 177. Oliver JD, Sibley GEM, Beckmann N, et al. F901318 represents a novel class of antifungal drug that inhibits dihydroorotate dehydrogenase. *Proc Natl Acad Sci U S A* 2016;113(45):12809–14.

178. du Pré S, Beckmann N, Almeida MC, et al. Effect of the novel antifungal drug F901318 (Olorofim) on growth and viability of *Aspergillus fumigatus*. *Antimicrob Agents Chemother* 2018;62(8):e00231.
179. Jørgensen KM, Astvad KMT, Hare RK, et al. EUCAST determination of Olorofim (F901318) susceptibility of mold species, method validation, and MICs. *Antimicrob Agents Chemother* 2018;62(8):e00487.
180. Kennedy T, Allen G, Steiner J, et al. Assessment of the duration of infusion on the tolerability and repeat dose pharmacokinetics of F901318 in healthy volunteers. In: Proceedings of the 27th European congress of clinical microbiology and infectious Diseases. 2017.
181. Kennedy T, Allen G, Steiner J, et al. An open-label study in healthy volunteers to evaluate the potential for cytochrome P450 3A4 inhibition by F901318 using oral midazolam as a probe. In: Proceedings of the 27th European congress of clinical microbiology and infectious Diseases. 2017.
182. Chen S, Rai NJ, Cunneen S, et al. A case of *Lomentospora prolificans* (LoPro) treated with the novel antifungal olorofim. In: Proceedings of the 30th European Congress of clinical Microbiology & infectious diseases. 2020.
183. Yen Tio S, Thrusky K, Ng G, et al. Olorofim for a case of severe disseminated *Lomentospora prolificans* infection. In: Proceedings of the 30th European congress of clinical microbiology & infectious Diseases. 2020.
184. Harvey E, Fitton L, Rex JH, et al. Successful use of the novel antifungal olorofim in the treatment of disseminated coccidiomycosis. In: Proceedings of the 30th European congress of clinical microbiology & infectious Diseases. 2020.
185. Tsukahara K, Hata K, Nakamoto K, et al. Medicinal genetics approach towards identifying the molecular target of a novel inhibitor of fungal cell wall assembly. *Mol Microbiol* 2003;48(4):1029–42.
186. Rivero-Menendez O, Cuenca-Estrella M, Alastruey-Izquierdo A. In vitro activity of APX001A against rare moulds using EUCAST and CLSI methodologies. *J Antimicrob Chemother* 2019;74(5):1295–9.
187. Zhao Y, Lee MH, Paderu P, et al. Significantly improved pharmacokinetics enhances in vivo efficacy of APX001 against echinocandin- and multidrug-resistant candida isolates in a mouse model of invasive candidiasis. *Antimicrob Agents Chemother* 2018;62(10):e00425.
188. Pappas PG, Kullberg BJ, Vazquez JA, et al. Clinical safety and efficacy of novel antifungal, fosmanogepix, in the treatment of Candidemia: results from a phase 2 proof of concept trial. In: Proceedings of IDWeek 2020. 2020.
189. FDA grants fast track status to amplyx pharmaceuticals for IV and oral formulations of fosmanogepix (APX001) for seven different indications. Available at: <https://amplyx.com/fda-grants-fast-track-status-to-amplyx-pharmaceuticals-for-iv-and-oral-formulations-of-fosmanogepix-apx001-for-seven-different-indications/>. Accessed February 20, 2021.